

THE ROLE OF PROSTAGLANDINS IN THE CENTRAL NERVOUS SYSTEM

◆1244

Leonhard S. Wolfe

Department of Neurology and Neurosurgery, McGill University, Montreal,
Quebec H3A 2B4, Canada

Flavio Coceani

Research Institute, The Hospital for Sick Children, Toronto,
Ontario M5G 1X8, Canada

INTRODUCTION

The prostaglandin system is implicated in physiological and pathological responses of most tissues of the body. The background and breadth of this subject are documented in many reviews and recent symposia (15, 25, 27, 28, 51, 108, 112, 114, 135). This review on the CNS covers advances in the past three years and concerns specifically the possible involvement of the prostaglandin system in the regulation of physiological and pathophysiological processes.

A wide range of stimuli (hormones, enzymes, trauma, inflammation, pyrogens, immune and allergic reactions, etc) activate a plasma membrane enzyme sequence in mammalian cells that leads to the rapid de novo synthesis of several prostaglandin types and in certain tissues thromboxanes as well. The biologically active compounds do not accumulate intracellularly and, therefore, under physiological conditions they occur only in trace amounts in tissues and most body fluids. Following formation, action and release, the compounds are rapidly converted by several enzymatic sequences to less active or inactive metabolites, which appear in blood and urine. Arachidonic acid, the predominant precursor unsaturated fatty acid in mammalian cells, becomes oxygenated to the prostaglandin endoperox-

ides and their products. Arachidonic acid must be released from a complex lipid precursor by deacylases before it can be transformed. The biosynthesis of prostaglandins and thromboxanes by central nervous tissue and factors that affect it have been reviewed recently (42, 135, 136). A new finding is that PGD_2 is formed in excess of $\text{PGF}_{2\alpha}$ by cerebral tissues of the rat (1).

CEREBROSPINAL FLUID

The existence of prostaglandin-like material in cerebrospinal fluid (CSF) of experimental animals has been recognized for some time (25, 135). Recent studies show CSF levels of $\text{PGF}_{2\alpha}$ in human subjects without neurological disease usually to be below 100 pg ml^{-1} (range $30\text{--}140 \text{ pg ml}^{-1}$) in cell-free fluid (55, 73, 137). Either PGE_2 is not detectable or it is present at the same low levels as $\text{PGF}_{2\alpha}$. Thromboxane B_2 is also a normal constituent (range $80\text{--}300 \text{ pg ml}^{-1}$), at least in the cat (F. Coceani, unpublished results). In contrast to most other tissues (89, 135), the brain has very low capacity either to take up or metabolize $\text{PGF}_{2\alpha}$ and PGE_2 to the 15-keto and 15-keto-13,14-dihydro metabolites. Consequently, prostaglandins normally produced endogenously are primarily cleared into the general circulation through choroidal and extra-choroidal transport mechanisms (16, 55).

Marked increases in CSF $\text{PGF}_{2\alpha}$ levels are found in patients with epilepsy, meningoencephalitis, hydrocephalus, and after surgical trauma; but levels are variable even in the same patient. Likewise, patients with vascular lesions, subarachnoid hemorrhage, and stroke also show significant albeit variable ($200\text{--}3000 \text{ pg ml}^{-1}$) increases in $\text{PGF}_{2\alpha}$ and PGE_2 levels (22, 55, 74, 137). Prostaglandins in the CSF may affect brain function directly or through local changes in the circulation.

CEREBRAL CIRCULATION

It is now well accepted that prostaglandins and thromboxanes contribute to vascular homeostasis through a direct action on smooth muscle in the vessel wall and, possibly, a modulation of muscle responses to neural and hormonal stimuli (27, 78, 125). The evidence supporting this concept is as follows: (a) Vessels are endowed with an enzyme system for the synthesis of primary prostaglandins and PGI_2 , the latter being the predominant prostaglandin. With one exception (umbilical artery), all vessels lack the thromboxane A_2 synthetic enzyme. (b) Prostaglandins and thromboxane A_2 exert potent and varied actions on vessels. While the action of the primary prostaglandins changes depending on the species and the vascular bed, PGI_2 and thromboxane A_2 are relaxant and constrictor agents, respectively, at all sites. Prostaglandin endoperoxides are also vasoactive, and

their action may be direct or mediated by the intramural formation of primary prostaglandins and PGI₂. (c) Indomethacin and other nonsteroidal anti-inflammatory drugs constrict or dilate vessels *in vitro* and *in vivo*.

Although these findings implicate intramural prostaglandins in the control of vascular tone, extramural prostaglandins may also be important, particularly under pathological conditions. Thromboxane A₂, which is released in great amounts from aggregating platelets, may gain access to muscle cells in the vessel wall and cause constriction. Furthermore, prostaglandins and thromboxane A₂ formed within the parenchyma of organs may act upon small resistance vessels.

The above scheme may also apply to the cerebral circulation. All primary prostaglandins and PGI₂ are formed in cerebral vessels (56, 133). Moreover, indomethacin reduces cerebral blood flow (99, 101), which implies that vessels are normally maintained in a relaxed state by a prostaglandin. The identity of the active compound is not known. However, evidence obtained in other vascular beds and the demonstration that PGE₂ and PGF_{2α} are both constrictors on cerebral vessels (100, 139) suggest that this compound is PGI₂. Thromboxane A₂, though not formed in cerebral vessels (56), is a potent constrictor (39). In fact, thromboxane A₂ is the most potent vasoconstrictor among agents acting on the cerebral circulation.

According to current ideas, prostaglandins and allied compounds, besides being involved in the normal control of cerebral blood flow, are also responsible for the hemodynamic changes occurring under certain pathological conditions—in particular, cerebral vasospasm (132). For example, thromboxane A₂, formed in damaged brain tissue or in aggregating platelets, is considered a prime determinant of the vasospasm-complicating thromboembolism and subarachnoid hemorrhage (39, 131). Thromboxane A₂ action may be complemented by that of the prostaglandins and other vasoactive agents (5-hydroxytryptamine) (2). Indeed, thrombin stimulates PGF_{2α} and PGE₂ synthesis when injected intrathecally (54). Brain ischemia following head injury is possibly another prostaglandin-mediated process. Prostaglandins, specifically PGE₂, have also been implicated in the pathogenesis of migraine (58, 131). PGE₂, a constrictor of intracranial vessels, dilates extracranial vessels (100); therefore, it may be involved both in the prodromal phase and in the headache phase of the migraine attack.

HYPOTHALAMIC FUNCTION

Prostaglandins have been implicated in several hypothalamic mechanisms. Only temperature regulation, water balance, and food intake are considered here. Involvement of prostaglandins in hypothalamo-adenohypophyseal function has been discussed in recent reviews (53, 68, 70, 110).

Temperature Regulation

The subject of hypothalamic transmitters involved in thermal homeostasis has been well covered recently (29, 42, 62, 77, 99, 107, 128, 140) and does not require further elaboration here. It is sufficient to say that three compounds, 5-hydroxytryptamine (5-HT), norepinephrine (NE), and acetylcholine (ACh) are generally assigned a key role in temperature regulation. According to most authors, body temperature is controlled through the opposing actions of 5-HT and NE on neurons in the anterior hypothalamic/preoptic region (AH/POA). These amines have species-specific signs of action while maintaining reciprocal effects. ACh is considered a transmitter in the temperature-raising pathway in all species. The extracellular concentration of ions within the posterior hypothalamus, and specifically the balance between sodium and calcium, may be an additional controlling factor. This ionic mechanism is thought to work in concert with the neurohumoral mechanism to determine the "set-point" around which body temperature is regulated.

The prostaglandins are a relatively recent addition to the field of thermoregulation. Interest in these compounds dates back to the early 1970s when it was found that PGE₁ was a potent pyretic agent (84) and that antipyretics blocked prostaglandin synthesis in various organs including brain (48, 112). These two findings implicated a prostaglandin in the genesis of fever. Research in this area developed actively and led to the demonstration that: (a) PGE₂, a normal constituent of hypothalamic tissue (70), is as potent as PGE₁ in producing fever (45-47, 57, 65, 76, 86, 93, 94, 102, 106, 121); moreover, both compounds are like pyrogens in that their action is not influenced by ambient temperature (57, 65, 121, 126); (b) PGE₂ acts upon neurons in the AH/POA that are also the main target for pyrogens (121, 126); (c) thermo-sensitive neurons in AH/POA respond in the same manner to PGE₂ and pyrogens (117); (d) PGE₂ fever, unlike pyrogen fever, does not abate following administration of antipyretics (24, 79, 84, 85); and (e) pyrogen fever is associated with elevated levels in the CSF of a prostaglandin with the biological and immunological properties of PGE₂ (34, 37, 43, 44, 83, 95). Collectively, these findings indicate that PGE₂ is well suited for being the "central messenger" of fever and specifically of pyrogen fever (cf 62, 80, 128). According to current knowledge, pyrogens from outside the body (*exogenous pyrogen*), and foremost among them bacterial endotoxin, as well as pathological conditions causing tissue inflammation and damage (e.g. infarction, malignancy) elicit the formation of a pyrogenic substance (*endogenous pyrogen*) in neutrophils and in cells of the reticuloendothelial system. The endogenous pyrogen, which is therefore a key intermediate in the sequence of events leading to fever, is then carried to the rostral region

of the hypothalamus by the circulation. Because the blood-brain barrier is seemingly impermeable to endogenous pyrogen (cf 80), and because prostaglandins are rapidly removed from the circulation (cf 124), one must assume that the vessel wall is the main site where pyrogen action is translated into increased prostaglandin synthesis. Consistent with this hypothesis is the notion that vessels, including cerebral vessels, are endowed with an active prostaglandin-generating system and that hypothalamic blood flow is increased during pyrogen fever (109). The latter finding implies activation of prostaglandin synthesis in the vessel wall. Alternatively, PGE₂ could be released from phagocytosing leucocytes sequestered in the capillary bed of AH/POA (128). Any pyrogen crossing the blood-brain barrier may stimulate prostaglandin synthesis in neural tissue (140). PGE₂, whether formed in the tissue of the AH/POA or from the vessels, acts at appropriate sites in the thermoregulatory pathways to elevate the "set-point" for temperature regulation, thus causing fever. Once its action is completed, PGE₂ is either inactivated enzymatically in situ or enters the extracellular fluid and CSF whence it is transported into the circulation. Interference with the latter mechanism results in enhancement of pyrogen effects (30).

Although the experimental evidence implicating PGE₂ in the pathogenesis of fever seems quite convincing, there have been reports contradicting the above scheme. In the monotreme, *Tachyglossus aculeatus* (Echidna), PGE₁ and PGE₂ are hypothermic agents, whereas endotoxin causes fever (12). Dissociation between pyrogen and PGE effects also occurs in the newborn lamb which, after appropriate sensitization, may develop fever in response to pyrogens but not in response to prostaglandins (103, 105). A similar phenomenon has been described in the adult animal following destruction of AH/POA (126). Potentially germane to these findings is the demonstration that prostaglandin antagonists block PGE₂ but not pyrogen fever (31).

Because pyrogen fever is susceptible to antipyretic treatment in the above experiments, a possible explanation for the inconsistencies could be that an arachidonic acid metabolite other than PGE₂ contributes to, or is the main determinant of pyrogen effects. Consistent with this is the finding that fever following administration of arachidonic acid, while abolished by antipyretics, is only partially blocked by prostaglandin antagonists (71). Theoretically, several compounds could have this role; however, available data limit the choice to two compounds, PGI₂ and thromboxane A₂, because prostaglandin endoperoxides and PGD₂ are inactive (41, 59), and PGF_{2α} is a pyretic agent but only in high doses (41, 47, 86). While no information is available on the central action of PGI₂, recent work showing that levels of thromboxane B₂ in the cerebrospinal fluid rise during pyrogen fever (F. Coceani, unpublished results) suggests that thromboxane A₂ might be the

hypothetical mediator. If so, it would not be a coincidence that intracranial bleeding, a condition in which AH/POA may be exposed to massive amounts of thromboxane A₂ formed in aggregating platelets, is commonly associated with fever (113).

Some findings suggest that fever may develop independently of the prostaglandin system. In the rabbit, salicylate at certain doses has little or no effect on the febrile response to pyrogen while it completely reverses the elevation in prostaglandin levels in the CSF (34). The question remains whether the dose of salicylate used was sufficient to block the synthesis of PGE₂ or any other pyrogenic derivative of arachidonic acid in the AH/POA. However, more cogent evidence against the involvement of the prostaglandin system in fever is afforded by recent work in the chick (4, 5) in which it was shown that PGEs are hyper- or hypothermic agents depending on the ambient temperature and that pyrogen fever is only marginally affected by indomethacin at a dose exceeding the therapeutic range (cf 48). Etiocolanolone fever in man, which is mediated by endogenous pyrogen (19), is also resistant to antipyretic treatment.

Summing up, a large body of evidence supports the existence of a "PGE₂ link" in the central action of endogenous pyrogen; but this prostaglandin may work in concert with another product, or more than one product, of arachidonic acid metabolism. Some forms of pyrogen fever, however, do not involve the prostaglandin system, and their central mechanism remains obscure.

Prostaglandins probably do not contribute to normal temperature regulation. Antipyretics, whether given systemically (24, 66, 79) or injected into the anterior hypothalamus (6, 35), produce little or no hypothermia in the afebrile animal, nor do they reverse the hyperthermia following cold stress (32, 104). Furthermore, prostaglandin levels in the CSF remain unchanged during thermoregulatory adjustments to cold or hot environments (21, 33). When present, hypothermic effects of antipyretics (cf 115) are ascribed to activation of the heat loss mechanism rather than to blockade of prostaglandin synthesis (79). Indeed, iontophoretically applied salicylate may stimulate warm-sensitive neurons in the AH/POA of the afebrile animal (13).

The intimate mechanism of prostaglandin action in producing fever remains a subject of speculation. It has been debated for some time (cf 28, 62) whether the prostaglandin and monoaminergic mechanisms are functionally interdependent, and this issue is far from being settled. In essence, two schemes have been proposed for linking the prostaglandins, specifically PGE₁ and PGE₂, to the monoamines. According to one (7), monoamine actions leading to elevation in body temperature are mediated in part by a prostaglandin. In support of this concept is the finding that 5-HT stimulates the release of PGEs from brain (64) and that epinephrine as well as 5-HT-

induced hyperthermia are suppressed by antipyretics (7, 69, 82, 85). However, the validity of results with the antipyretics has been questioned (36, 82), moreover, this hypothesis is not easily reconciled with the notion that monoamine effects (17, 62), unlike prostaglandin effects (57, 65, 121, 126), are affected by ambient temperature. Alternatively, it has been suggested on the basis of work with specific monoamine depletors and antagonists that monoamines are intermediates in the action of prostaglandins on temperature-raising mechanisms. Again, no firm conclusion can be drawn from these studies (cf 28) because positive results in one species [rabbit (20, 66, 72)] contrast with inconsistent results in another [cat (82, 126)]. Furthermore, an explanation for the constancy of prostaglandin effects at different ambient temperatures must be provided before accepting this hypothesis. Equally controversial is the question of the role of cyclic nucleotides in PGE fever. While findings in the rabbit suggest that cyclic AMP is a central mediator of PGE fever (138), findings in the cat argue against this idea (81).

Control of Body Water

The homeostatic regulation of body water content is dependent on the concerted action of two brain mechanisms, namely, the function of a "thirst sensor" possibly located in the subformal organ (40) and other circumventricular organs (97), and the secretion of antidiuretic hormone (ADH). Both mechanisms are under the direct control of angiotensin II (40, 118) and may also be influenced by the prostaglandins. Angiotensin, whether formed in situ or blood-borne, stimulates thirst and the formation of ADH. The latter action is exerted on the synthesis [supraoptic and paraventricular neurons (87)] and release [neurohypophysis (50, 63)] of the hormone. When injected into the common carotid artery or the cerebral ventricles, PGE₁ and PGE₂ mimic angiotensin in stimulating ADH release (75, 129). Moreover, these prostaglandins share with angiotensin a dual site of action (50, 75, 129). Prostaglandin action on thirst mechanisms is a subject of controversy. While it has been reported that PGE₁ and PGE₂ (but not PGF_{2α}) antagonize the dipsogenic effect of angiotensin in the rat (40, 91), the same compounds have opposite effects in the goat (3, 75). This discrepancy is unlikely due to the dose of prostaglandin used (40, 75), because the sign of responses in the rat remained the same over a wide range of doses. Differences may reflect a genuine species variation, the significance of which is not known.

Because low doses of PGE₁ and PGE₂ given by the ventricular route affect water balance without altering thermoregulatory neurons, responses are possibly indicative of a physiological process. This applies specifically to PGE₂, which is present in brain. Future experiments employing blockers

of prostaglandin synthesis may confirm this point. Regardless of whether responses are physiological or pharmacological, the mechanism of prostaglandin action remains to be elucidated. Prostaglandins and angiotensin, which are both vasoactive agents, may act, or interact, on blood vessels supplying target neurons in the subfornical organ and the hypothalamus. Alternatively, their action may be exerted directly on neurons. Indeed, subfornical and supraoptic neurons respond to iontophoretically applied angiotensin (92, 96). It is still a question whether the same holds true with the prostaglandins.

Regulation of Food Intake

It is generally assumed that food intake is primarily controlled through the opposing action of two neuronal systems located in the hypothalamus: a lateral system signalling the urge for food ("feeding center"), and a ventromedial system suppressing food intake ("satiety center") (60). Several neurohumoral agents, including the prostaglandins, have been implicated in the function of these neurons (cf 8). When given systemically, various prostaglandin types, including PGE₁, PGE₂, and PGF_{2α}, inhibit food intake without overtly affecting behavior, body temperature, and water intake (116). PGE₁ is also effective when injected into the hypothalamus; however, its site of action varies with the species. While in the rat responsive sites are located in the anterior commissure region and the lateral hypothalamus (11), in the ewe they are located in the anterior and medial hypothalamus (10). Furthermore, in the ewe PGE₁ may also stimulate feeding (10). Prostaglandin effects occur in both food-deprived and satiated animals (38), which implies a central action for these compounds.

Although these findings implicate the prostaglandins in the hypothalamic control of energy balance, some facts are inconsistent with this possibility. PGE₂, even though it suppresses feeding by the systemic route, has no effect on sites in the hypothalamus that are sensitive to PGE₁ (10). Moreover, effective doses of PGE₁ by the intrahypothalamic route are in the microgram range (10, 11, 127, 134), indicating a pharmacological rather than a physiological action. Another difficulty in accepting this idea arises from the fact that distribution of prostaglandin-sensitive sites in the hypothalamus is species-specific in spite of a seemingly constant organization of the neuronal systems controlling feeding behavior (9).

INTERACTION WITH CYCLIC NUCLEOTIDES AND NEUROTRANSMITTERS

A large body of evidence suggests that cyclic nucleotides, and particularly adenosine 3',5'-monophosphate (cyclic AMP), play a role in peripheral and

central synapses (52, 67, 90). In the CNS, cyclic AMP has been implicated in the mediation of postsynaptic effects of several neurotransmitters (52, 67, 90); however, evidence of such a role is strongest in the case of dopaminergic synapses on caudate neurons (120) and β -adrenergic synapses between fibers originating in the locus coeruleus (LC) and cells in the cerebellum (Purkinje cells) and the hippocampus (pyramidal cells) (18). Some data also suggest that guanosine 3',5'-monophosphate (cyclic GMP) is involved in the muscarinic actions of ACh (52, 88, 90, 118). In fact, it has been proposed that cyclic AMP and cyclic GMP have a reciprocal function in the regulation of neuronal activity (123).

Prostaglandins may also interact with the cyclic nucleotides. Findings in cerebellar Purkinje cells afford a model of some of their possible functions in synaptic events. In brief, it is proposed (cf 28; 18, 90) that NE released from LC fibers impinging upon Purkinje cells triggers the postsynaptic formation of cyclic AMP, which in turn causes an appropriate change in membrane potential (i.e. hyperpolarization) through the phosphorylation of specific membrane proteins. The same model assumes that PGE_2 , formed in response to NE or cyclic AMP action, modulates the synaptic process by inhibiting the synthesis of cyclic AMP. A similar sequence of events is thought to occur in other noradrenergic synapses (90), whereas prostaglandins are assigned a stimulatory action on the cyclic AMP-generating system in dopaminergic synapses (120).

Although supported by findings in peripheral synapses (52), the above scheme has been challenged on various grounds. The idea that cyclic AMP is an essential intermediate in the postsynaptic action of NE has been questioned, and the points of contention are discussed in several reviews (28, 90, 98). Furthermore, different investigators (cf 23, 28, 122) have been unable to confirm at several sites in the CNS (cerebral cortex, hypothalamus and brain stem in mammals, and spinal cord in the frog) that E-type prostaglandins modify neuronal responses to the monoamines. Negative results with spinal neurons (23) are particularly significant because the amphibian CNS, unlike the mammalian CNS, is endowed with an active enzyme system for prostaglandin inactivation (26; F. Coceani, unpublished results) and would, therefore, seem to be a well-suited site for prostaglandin involvement in synaptic events. It is conceivable, however, that prostaglandins may influence postsynaptic actions of the monoamines only in certain neuronal systems. Future work must take into consideration effects of the endoperoxides and thromboxanes, which may turn out to be more important endogenous modulators of adenylate cyclase than the primary prostaglandins.

A separate line of investigation suggests that PGE_2 may modulate noradrenergic and dopaminergic transmission through inhibition of transmitter

release. However, this concept is based on work in peripheral synapses (61), whereas findings in the CNS are negative (130) or inconsistent (14, 111, 119).

CONCLUSIONS

Facts

Central nervous tissue has a complete system for the biosynthesis of prostaglandins and thromboxanes. The prostaglandin system is either directly or indirectly connected with neuronal activity, and its possible function is best documented in the case of hypothalamic homeostatic mechanisms. Cerebral blood vessels synthesize prostaglandins, which likely contribute to normal hemodynamics. There is compelling evidence that prostaglandins are formed at multiple sites in the CNS, both neural and non-neural, and interact in a varied manner in physiological and pathological situations.

Outstanding Issues

The activity and control of synthetic enzymes in neural and non-neural constituents of CNS; the identity of compounds active at various sites; the role of $\text{PGF}_{2\alpha}$ and PGD_2 , which are relatively inactive in spite of being formed in excess of PGE_2 ; the likelihood of prostaglandin-degrading enzymes being confined to certain neuronal types and the importance of such enzymes in the termination of prostaglandin effects; and the specific involvement of prostaglandins in synaptic events are the outstanding issues.

Prospectives

Most of the outstanding issues will be hard to resolve because of limitations in assay methodology, the multitude of compounds to be assayed, the potential for new compounds or pathways in the metabolism of arachidonic acid, and difficulties in following the time-sequence of biosynthetic events in vivo. In spite of these problems, refinements in assay methods and the development of new and more selective blockers of arachidonic acid metabolism should afford a better knowledge of the functional organization of the prostaglandin system in the CNS. Advances in this field should also have an impact in the clinic—particularly in the management of neurological diseases in which the neural deficit follows a vascular insult.

ACKNOWLEDGMENTS

Work of the authors of this review was supported by the Medical Research Council of Canada.

Literature Cited

1. Abdel-Halim, M. S., Hamberg, M., Sjöquist, B., Ånggård, E. 1977. Identification of prostaglandin D_2 as a major prostaglandin in homogenates of rat brain. *Prostaglandins* 14:633-43
2. Allen, G. S., Gross, C. J., French, L. A., Chou, S. N. 1976. Cerebral arterial spasm: *In vitro* contractile activity of vasoactive agents including human CSF on human basilar and anterior cerebral arteries. *J. Neurosurg.* 44:594-600
3. Andersson, B., Leksell, L. G. 1975. Effects on fluid balance of intraventricular infusions of prostaglandin E_1 . *Acta Physiol. Scand.* 93:286-88
4. Artunkal, A. A., Marley, E., Stephenson, J. D. 1977. Some effects of prostaglandins E_1 and E_2 and of endotoxin injected into the hypothalamus of young chicks: dissociation between endotoxin fever and the effects of prostaglandins. *Br. J. Pharmacol.* 61:39-46
5. Artunkal, A. A., Marley, E., Stephenson, J. D. 1977. Some effects of intravenous prostaglandin E_1 and endotoxin in young chickens. *Br. J. Pharmacol.* 61:29-37
6. Avery, D. D., Penn, P. E. 1974. Blockage of pyrogen induced fever by intrahypothalamic injections of salicylate in the rat. *Neuropharmacology* 13: 1179-85
7. Avery, D. D., Penn, P. E. 1976. Interaction of salicylate and body temperature changes caused by injections of neurohumours into the anterior hypothalamus: possible mechanisms of salicylate antipyresis. *Neuropharmacology* 15:433-38
8. Baile, C. A. 1974. Putative neurotransmitters in the hypothalamus and feeding. *Fed. Proc.* 33:1166-75
9. Baile, C. A., Forbes, J. M. 1974. Control of feed intake and regulation of energy balance in ruminants. *Physiol. Rev.* 54:160-214
10. Baile, C. A., Martin, F. H., Forbes, J. M., Webb, R. L., Kingsbury, W. 1974. Intrahypothalamic injections of prostaglandins and prostaglandin antagonists and feeding in sheep. *J. Dairy Sci.* 57: 81-88
11. Baile, C. A., Simpson, C. W., Bean, S. M., McLaughlin, C. L., Jacobs, H. L. 1973. Prostaglandins and food intake of rats: a component of energy balance regulation? *Physiol. Behav.* 10:1077-85
12. Baird, J. A., Hales, J. R. S., Lang, W. J. 1974. Thermoregulatory responses to the injection of monoamines, acetylcholine and prostaglandins into a lateral ventricle of the echidna. *J. Physiol. London* 236:539-48
13. Beckman, A. L., Rozkowska-Ruttimann, E. 1974. Hypothalamic and septal neuronal responses to iontophoretic application of salicylate in rats. *Neuropharmacology* 13:393-98
14. Bergström, S., Farnebo, L.-O., Fuxe, K. 1973. Effect of prostaglandin E_2 on central and peripheral catecholamine neurons. *Eur. J. Pharmacol.* 21:362-368
15. Berti, F., Samuelsson, B., Velo, G. P., eds. 1976. *Prostaglandins and Thromboxanes*. NATO Advanced Study Institute Series A., Vol. 13. New York: Plenum. 449 pp.
16. Bito, L. Z., Davson, H., Hollingsworth, J. R. 1976. Facilitated transport of prostaglandins across the blood-cerebrospinal fluid and blood-brain barriers. *J. Physiol. London* 256:273-85
17. Bligh, J., Cottle, W. H., Maskrey, M. 1971. Influence of ambient temperature on the thermoregulatory responses to 5-hydroxytryptamine, noradrenaline and acetylcholine injected into the lateral cerebral ventricles of sheep, goats and rabbits. *J. Physiol. London* 212: 377-92
18. Bloom, F. E., Siggins, G. R., Hoffer, B. J., Segal, M., Oliver, A. P. 1975. Cyclic nucleotides in the central synaptic actions of catecholamines. In *Advances in Cyclic Nucleotide Research*, ed. G. I. Drummond, P. Greengard, G. A. Robinson, 5:603-18. New York: Raven. 872 pp.
19. Bodel, P., Dillard, M. 1968. Studies on steroid fever. I. Production of leukocyte pyrogen in vitro by ethiocholanolone. *J. Clin. Invest.* 47:107-17
20. Borsook, D., Laburn, H. P., Rosendorff, C., Willies, G. H., Woolf, C. J. 1977. A dissociation between temperature regulation and fever in the rabbit. *J. Physiol. London* 266:423-33
21. Cammock, S., Dascombe, M. J., Milton, A. S. 1976. Prostaglandins in thermoregulation. See Ref. 114, 1:375-80
22. Carasso, R. L., Vardi, J., Rabay, J. M., Zor, U., Streifler, M. 1977. Measurement of prostaglandin E_2 in cerebrospinal fluid in patients suffering from stroke. *J. Neurol. Psychiatr.* 40:967-69
23. Caulford, P. G., Cocceani, F. 1977. Microiontophoresis of 5-hydroxytryptamine, epinephrine and prostaglandin E_1 on spinal neurons in the frog. *Can. J. Physiol. Pharmacol.* 55:293-300
24. Clark, W. G., Cumby, H. R. 1975. The

- antipyretic effect of indomethacin. *J. Physiol. London* 248:625-38
25. Coceani, F. 1974. Prostaglandins and the central nervous system. *Arch. Intern. Med.* 133:119-29
 26. Coceani, F. 1978. Studies of the prostaglandins in the frog spinal cord. In *Iontophoresis and Transmitter Mechanisms in the Mammalian Central Nervous System*, ed. R. W. Ryall, J. S. Kelly, pp. 456-58. New York: Elsevier. 494 pp.
 27. Coceani, F., Olley, P. M., eds. 1978. Prostaglandins and perinatal medicine. In *Advances in Prostaglandin and Thromboxane Research*, Vol. 4. New York: Raven. 412 pp.
 28. Coceani, F., Pace-Asciak, C. R. 1976. Prostaglandins and the central nervous system. In *Prostaglandins: Physiological, Pharmacological and Pathological Aspects*, ed. S. M. M. Karim, pp. 1-36. Lancaster, Pa.: MTP Press. 367 pp.
 29. Cooper, K. E., Lomax, P., Schönbaum, E., eds. 1977. *Drugs, Biogenic Amines and Body Temperature*. Basel: Karger. 283 pp.
 30. Cooper, K. E., Veale, W. L. 1972. The effect of injecting an inert oil into the cerebral ventricular system upon fever produced by intravenous leucocyte pyrogen. *Can. J. Physiol. Pharmacol.* 50:1066-71
 31. Cranston, W. I., Duff, G. W., Hellon, R. F., Mitchell, D., Townsend, Y. 1976. Evidence that brain prostaglandin synthesis is not essential in fever. *J. Physiol. London* 259:239-49
 32. Cranston, W. I., Hellon, R. F., Luff, R. H., Rawlins, M. D., Rosendorff, C. 1970. Observations on the mechanism of salicylate-induced antipyresis. *J. Physiol. London* 210:593-600
 33. Cranston, W. I., Hellon, R. F., Mitchell, D. 1975. Is brain prostaglandin synthesis involved in responses to cold? *J. Physiol. London* 249:425-34
 34. Cranston, W. I., Hellon, R. F., Mitchell, D. 1975. A dissociation between fever and prostaglandin concentration in cerebrospinal fluid. *J. Physiol. London* 253:583-92
 35. Cranston, W. I., Rawlins, M. D. 1972. Effects of intracerebral microinjection of sodium salicylate on temperature regulation in the rabbit. *J. Physiol. London* 222:257-66
 36. Dey, P. K., Feldberg, W., Gupta, K. P., Milton, A. S., Wendlandt, S. 1974. Further studies on the role of prostaglandins in fever. *J. Physiol. London* 241:629-46
 37. Dey, P. K., Feldberg, W., Wendlandt, S. 1974. Lipid A and prostaglandin. *J. Physiol. London* 239:102-3P
 38. Doggett, N. S., Jawaharlal, K. 1977. Some observations on the anorectic activity of prostaglandin F_{2α}. *Br. J. Pharmacol.* 60:409-15
 39. Ellis, E. F., Nies, A. S., Oates, J. A. 1977. Cerebral arterial smooth muscle contraction by thromboxane A₂. *Stroke* 8:480-83
 40. Epstein, A. N. 1978. The neuroendocrinology of thirst and salt appetite. In *Frontiers in Neuroendocrinology*, ed. W. F. Ganong, L. Martini, 5:101-34. New York: Raven. 399 pp.
 41. Ewen, L., Milton, A. S., Smith S. 1976. Effects of prostaglandin F_{2α} and prostaglandin D₂ on the body temperature of conscious cats. *J. Physiol. London* 258:121-22P
 42. Feldberg, W. 1975. Body temperature and fever: changes in our views during the last decade. *Proc. R. Soc. London Ser. B* 191:199-229
 43. Feldberg, W., Gupta, K. P. 1973. Pyrogen fever and prostaglandin-like activity in cerebrospinal fluid. *J. Physiol. London* 228:41-53
 44. Feldberg, W., Gupta, K. P., Milton, A. S., Wendlandt, S. 1973. Effect of pyrogen and antipyretics on prostaglandin activity in cisternal c.s.f. of unanaesthetized cats. *J. Physiol. London* 234:279-303
 45. Feldberg, W., Saxena, P. N. 1971. Fever produced by prostaglandin E₁. *J. Physiol. London* 217:547-56
 46. Feldberg, W., Saxena, P. N. 1971. Further studies on prostaglandin E₁ fever in cats. *J. Physiol. London* 219:739-45
 47. Feldberg, W., Saxena, P. N. 1975. Prostaglandins, endotoxin and lipid A on body temperature in rats. *J. Physiol. London* 249:601-15
 48. Flower, R. J. 1974. Drugs which inhibit prostaglandin synthesis. *Pharmacol. Rev.* 26:33-67
 49. Fumagalli, R., Folco, G. C., Longiave, D. 1976. Influence of prostaglandins on central functions. See Ref. 15, pp. 383-421
 50. Gagnon, D. J., Cousineau, D., Boucher, P. J. 1973. Release of vasopressin by angiotensin II and prostaglandin E₂ from the rat neurohypophysis in vitro. *Life Sci.* 12:487-97
 51. Galli, C., Galli, G., Porcellati, G., eds. 1978. Phospholipases and prostaglandins. In *Advances in Prostaglandin and Thromboxane Research*, Vol. 3. New York: Raven. 206 pp.

52. Grøengard, P. 1976. Possible role for cyclic nucleotides and phosphorylated membrane proteins in post-synaptic actions of neurotransmitters. *Nature* 260:101-8
53. Hafsi, H. D., Haynes, N. B. 1977. Prostaglandins and pituitary hormone secretion. In *Prostaglandins and Therapeutics*, ed. D. K. Silver, 3(3):3-4. Kalamazoo, Mich.: The Upjohn Company. 4 pp.
54. Hagen, A. A., Gerber, J. N., Sweeley, C. C., White, R. P., Robertson, J. T. 1977. Pleocytosis and elevation of prostaglandin F_{2a} and E_2 in cerebrospinal fluid following intracisternal injection of thrombin. *Stroke* 8:236-38
55. Hagen, A. A., Gerber, J. N., Sweeley, C. C., White, R. P., Robertson, J. T. 1977. Levels and disappearance of prostaglandin F_{2a} in cerebral spinal fluid: a clinical and experimental study. *Stroke* 8:672-75
56. Hagen, A. A., White, R. P., Terragno, N. A., Robertson, J. T. 1978. Synthesis of prostaglandins (PGs) by bovine cerebral arteries. *Fed. Proc.* 37:384
57. Hales, J. R. S., Bennett, J. W., Baird, J. A., Fawcett, A. S. 1973. Thermoregulatory effects of prostaglandins E_1 , E_2 , F_{1a} and F_{2a} in the sheep. *Pfleugers Arch.* 339:125-33
58. Harper, A. M., McCulloch, J., MacKenzie, E. T., Pickard, J. D. 1977. Migraine and the blood-brain barrier. *Lancet* 1:1034-36
59. Hawkins, M., Lipton, J. M. 1977. Analogs of endoperoxide precursors of prostaglandins: failure to affect body temperature when injected into primary and secondary central temperature controls. *Prostaglandins* 13:209-18
60. Hayward, J. N. 1977. Functional and morphological aspects of hypothalamic neurons. *Physiol. Rev.* 57:574-658
61. Hedqvist, P. 1973. Autonomic neurotransmission. See Ref. 108, 1:101-31
62. Hellon, R. F. 1974. Monoamines, pyrogens and cations: their actions on central control of body temperature. *Pharmacol. Rev.* 26:289-321
63. Hisada, S., Fujimoto, S., Kamiya, T., Endo, Y., Tsumahima, H. 1977. Antidiuresis of centrally administered amines and peptides and release of antidiuretic hormone from isolated rat neurohypophysis. *Jpn. J. Pharmacol.* 27:153-61
64. Holmes, S. W. 1970. The spontaneous release of prostaglandins into the cerebral ventricles of the dog and the effect of external factors on this release. *Br. J. Pharmacol.* 38:653-58
65. Hori, T., Harada, Y. 1974. The effects of ambient and hypothalamic temperatures on the hyperthermic responses to prostaglandins E_1 and E_2 . *Pfleugers Arch.* 350:123-34
66. Kandasamy, B., Girault, J.-M., Jacob, J. 1975. Central effects of a purified bacterial pyrogen, prostaglandin E_1 and biogenic amines on the temperature in the awake rabbit. See Ref. 77, pp. 124-32
67. Keabadian, J. W. 1977. Biochemical regulation and physiological significance of cyclic nucleotides in the nervous system. In *Advances in Cyclic Nucleotide Research*, ed. P. Greengard, G. A. Robinson, 8:421-508. New York: Raven. 582 pp.
68. Kenimer, J. G., Goldberg, V., Blecker, M. 1977. The endocrine system: interactions of prostaglandins with adenylyl cyclase-cyclic AMP systems. See Ref. 108, 3:77-108
69. Komiskey, H. L., Rudy, T. A. 1975. The involvement of methysergide-sensitive receptors and prostaglandins in the hyperthermia evoked by 5-HT in the cat. *Res. Commun. Chem. Path. Pharmacol.* 11:195-208
70. Labrie, F., Pelletier, G., Borgeat, P., Drouin, J., Ferland, L., Belanger, A. 1976. Mode of action of hypothalamic regulatory hormones in the adeno-hypophysis. In *Frontiers in Neuroendocrinology*, ed. L. Martini, W. F. Ganong, 4:63-93. New York: Raven. 294 pp.
71. Laburn, H., Mitchell, D., Rosendorff, C. 1977. Effects of prostaglandin antagonism on sodium arachidonate fever in rabbits. *J. Physiol. London* 267:559-70
72. Laburn, H., Woolf, C. J., Willies, G. H., Rosendorff, C. 1975. Pyrogen and prostaglandin fever in the rabbit. II. Effects of noradrenaline depletion and adrenergic receptor blockade. *Neuropharmacology* 14:405-11
73. Landau, I. S., Young, C. W. 1977. Measurement of prostaglandin F_{2a} levels in cerebrospinal fluid of febrile and afebrile patients with advanced cancer. *Prostaglandins* 14:343-53
74. Latorre, E., Patrono, C., Fortuna, A., Grossi-Belloni, D. 1974. Role of prostaglandin F_{2a} in human cerebral vasospasm. *J. Neurosurg.* 41:293-99
75. Leksell, L. G. 1976. Influence of prostaglandin E_1 on cerebral mechanisms involved in the control of fluid balance. *Acta Physiol. Scand.* 98:85-93
76. Lipton, J. M., Fossler, D. E. 1974. Fever produced in the squirrel monkey by

- intravenous and intracerebral endotoxin. *Am. J. Physiol.* 226:1022-27
77. Lomax, P., Schönbaum, E., Jacob, J., eds. 1975. *Temperature Regulation and Drug Action*. Basel: Karger. 405 pp.
 78. Malik, K. U. 1978. Prostaglandins—modulation of adrenergic nervous system. *Fed. Proc.* 37:203-7
 79. Milton, A. S. 1973. Prostaglandin E₁ and endotoxin fever, and the effects of aspirin, indomethacin, and 4-acetamidophenol. *Adv. Biosci.* 9:495-500
 80. Milton, A. S. 1976. Modern views on the pathogenesis of fever and the mode of action of antipyretic drugs. *J. Pharm. Pharmacol.* 28:393-99
 81. Milton, A. S., Dascombe, M. J. 1977. Cyclic nucleotides in thermoregulation and fever. In *Drugs, Biogenic Amines, and Body Temperature*, ed. K. E. Cooper, P. Lomax, E. Schönbaum, pp. 129-35. Basel: Karger. 283 pp.
 82. Milton, A. S., Harvey, C. A. 1975. Prostaglandins and monoamines in fever. See Ref. 77, pp. 133-42
 83. Milton, A. S., Smith, S., Tomkins, K. B. 1977. Levels of prostaglandin F and E in cerebrospinal fluid of cats during pyrogeninduced fever. *Br. J. Pharmacol.* 59:447-48P
 84. Milton, A. S., Wendlandt, S. 1970. A possible role for prostaglandin E₁ as a modulator for temperature regulation in the central nervous system of the cat. *J. Physiol. London* 207:76-77P
 85. Milton, A. S., Wendlandt, S. 1971. The effects of 4-acetamidophenol (paracetamol) on the temperature response of the conscious rat to the intracerebral injection of prostaglandin E₁, adrenaline and pyrogen. *J. Physiol. London* 217:33-34P
 86. Milton, A. S., Wendlandt, S. 1971. Effects on body temperature of prostaglandins of the A, E, and F series on injection into the third ventricle of unanaesthetized cats and rabbits. *J. Physiol. London* 218:325-36
 87. Mouw, D., Bonjour, J. P., Malvin, R. L., Vander, A. 1971. Central action of angiotensin in stimulating ADH release. *Am. J. Physiol.* 220:239-42
 88. Nahorski, S. R., Pratt, C. N. F. W., Rogers, K. J. 1976. Increased cerebral cyclic GMP concentration induced by muscarinic cholinergic agonists and prostaglandin F_{2a}. *Br. J. Pharmacol.* 57:445-46P
 89. Nakano, J., Prancan, A. V., Moore, S. E. 1972. Metabolism of prostaglandin E₁ in cerebral cortex and cerebellum of the dog and rat. *Brain Res.* 39:545-48
 90. Nathanson, J. A. 1977. Cyclic nucleotides and the nervous system function. *Physiol. Rev.* 57:157-256
 91. Nicolaïdes, S., Fitzsimmons, J. T. 1975. La dépendance de la prise d'eau induite par l'angiotensine II envers la fonction vasomotrice cérébrale locale chez le Rat. *C. R. Acad. Sci. Paris Ser. D* 281:1417-20
 92. Nicoll, R. A., Barker, J. L. 1971. Excitation of supraoptic neurosecretory cells by angiotensin II. *Nature* 233:172-74
 93. Nisticó, G., Marley, E. 1973. Central effects of prostaglandin E₁ in adult fowls. *Neuropharmacology* 12:1009-16
 94. Nisticó, G., Marley, E. 1976. Central effects of prostaglandins E₂, A₁ and F_{2a} in adult fowls. *Neuropharmacology* 15:737-41
 95. Philipp-Dormston, W. K., Siegert, R. 1974. Prostaglandins of the E and F series in rabbit cerebrospinal fluid during fever induced by Newcastle Disease Virus, *E. coli*—endotoxin, or endogenous pyrogen. *Med. Microbiol. Immunol.* 159:279-84
 96. Phillips, M. I., Felix, D. 1976. Specific angiotensin II receptive neurons in the cat subfornical organ. *Brain Res.* 109:531-40
 97. Phillips, M. I., Hoffman, W. E. 1977. Sensitive sites in the brain for the blood pressure and drinking responses to angiotensin II. In *Central Actions of Angiotensin and Related Hormones*, ed. J. P. Buckley, G. Ferrario, pp. 325-56. New York: Pergamon. 580 pp.
 98. Phillis, J. W. 1977. The role of cyclic nucleotides in the CNS. *Can. J. Neurol. Sci.* 4:151-95
 99. Pickard, J. D., MacDonell, L. A., MacKenzie, E. T., Harper, A. M. 1977. Response of the cerebral circulation in baboons to changing perfusion pressure after indomethacin. *Circ. Res.* 40:198-203
 100. Pickard, J. D., MacDonell, L. A., MacKenzie, E. T., Harper, A. M. 1977. Prostaglandin-induced effects in the primate cerebral circulation. *Eur. J. Pharmacol.* 43:343-51
 101. Pickard, J. D., MacKenzie, E. T. 1973. Inhibition of prostaglandin synthesis and the response of baboon cerebral circulation to carbon dioxide. *Nature* 245:187-88
 102. Pittman, Q. J., Veale, W. L., Cockeram, A. W., Cooper, K. E. 1976. Changes in body temperature produced by prostaglandins and pyrogens in the chicken. *Am. J. Physiol.* 230:1284-87

103. Pittman, Q. J., Veale, W. L., Cooper, K. E. 1975. Temperature responses of lambs after centrally injected prostaglandins and pyrogens. *Am. J. Physiol.* 228:1034-38
104. Pittman, Q. J., Veale, W. L., Cooper, K. E. 1976. Observations on the effect of salicylate in fever and the regulation of body temperature against cold. *Can. J. Physiol. Pharmacol.* 54:101-6
105. Pittman, Q. J., Veale, W. L., Cooper, K. E. 1977. Effect of prostaglandin, pyrogen and noradrenaline injected into the hypothalamus, on thermoregulation in newborn lambs. *Brain Res.* 128:473-83
106. Potts, W. J., East, P. F. 1972. Effects of prostaglandin E₂ on the body temperature of conscious rats and cats. *Arch. Int. Pharmacodyn. Ther.* 197:31-36
107. Preston, E., Schönbaum, E. 1976. Monoaminergic mechanisms in thermoregulation. In *Brain Dysfunction in Infantile Febrile Convulsions*, ed. M. A. B. Brazier, F. Coceani, pp. 75-87. New York: Raven. 370 pp.
108. Ramwell, P. W., ed. 1973, 1974, 1977. *The Prostaglandins*, Vols. 1, 2, 3. New York: Plenum. 400 pp.; 350 pp.; 359 pp.
109. Rawlins, M. D., Luff, R. H., Cranston, W. I. 1973. Regional brain salicylate concentrations in afebrile and febrile rabbits. *Biochem. Pharmacol.* 22: 2639-42
110. Roberts, J. S., Carlson, J. C., McCracken, J. A. 1976. Prostaglandin F_{2a} production by the brain and its role in LH secretion. See Ref. 114, 2:609-19
111. Roberts, P. J., Hillier, K. 1976. Facilitation of noradrenaline release from rat brain synaptosomes by prostaglandin. *Brain Res.* 112:425-28
112. Robinson, H. J., Vane, J. R., eds. 1974. *Prostaglandin Synthetase Inhibitors*. New York: Raven. 395 pp.
113. Rudy, T. A., Westergaard, J. L., Yaksh, T. L. 1978. Hyperthermia produced by simulated intraventricular hemorrhage in the cat. *Exp. Neurol.* 58:296-310
114. Samuelsson, B., Paoletti, R., eds. 1976. *Advances in Prostaglandin and Thromboxane Research*, Vols. 1, 2. New York: Raven. 1028 pp.
115. Satinoff, E. 1972. Salicylate: action on normal body temperature in rats. *Science* 176:532-33
116. Scaramuzzi, O. E., Baile, C. A., Mayer, J. 1971. Prostaglandins and food intake of rats. *Experientia* 27:256-57
117. Schoener, E. P., Wang, S. C. 1976. Effects of locally administered prostaglandin E₁ on anterior hypothalamic neurons. *Brain Res.* 117:157-62
118. Severs, W. B., Daniels-Severs, A. E. 1973. Effects of angiotensin on the central nervous system. *Pharmacol. Rev.* 25:415-49
119. Shenoy, A., Ziance, R. 1978. Modulation of ³H-norepinephrine release in rat cerebral cortex by prostaglandin E₂ and autonomic drugs. *Fed. Proc.* 37:688
120. Siggins, G. R., Hoffer, B. J., Bloom, F. E., Understedt, U. 1976. Cytochemical and electrophysiological studies of dopamine in the caudate nucleus. *Res. Publ. Assoc. Res. Nerv. Ment. Dis.* 55: 227-48
121. Stitt, J. T. 1973. Prostaglandin E₁ fever induced in rabbits. *J. Physiol. London* 232:163-79
122. Stitt, J. T., Hardy, J. D. 1975. Microelectrophoresis of PGE₁ onto single units in the rabbit hypothalamus. *Am. J. Physiol.* 229:240-45
123. Stone, T. W., Taylor, D. A., Bloom, F. E. 1975. Cyclic AMP and cyclic GMP may mediate opposite neuronal responses in the rat cerebral cortex. *Science* 187:845-47
124. Vane, J. R. 1969. The release and fate of vasoactive hormones in the circulation. *Br. J. Pharmacol.* 35:209-42
125. Vane, J. R., McGiff, J. C. 1975. Possible contribution of endogenous prostaglandins to the control of blood pressure. *Circ. Res.* 36, 37: (Suppl.) I68-75
126. Veale, W. L., Cooper, K. E. 1975. Comparison of sites of action of prostaglandin E and leucocyte pyrogen in brain. See Ref. 77, 218-26
127. Veale, W. L., Cooper, K. E., Malkinson, T. 1976. Temperature and feeding responses in the unanaesthetized cat following injections of prostaglandin E into the hypothalamus. *Canada Physiol.* 7:64
128. Veale, W. L., Cooper, K. E., Pittman, Q. J. 1977. Role of prostaglandins in fever and temperature regulation. See Ref. 108, 3:145-62
129. Vilhardt, H., Hedqvist, P. 1970. A possible role of prostaglandin E₂ in the regulation of vasopressin secretion in rats. *Life Sci.* 9:825-30
130. VonVoigtlander, P. F. 1976. In vivo dopamine release and prostaglandin E₂. *Res. Comm. Chem. Pathol. Pharmacol.* 14:431-36
131. Welch, K. M. A., Spira, P. J., Knowles, L., Ance, J. W. 1974. Effects of prostaglandins in the internal and external carotid blood flow in the monkey. *Neurology* 24:705-10
132. White, R. P., Hagen, A. A., Morgan, H., Dawson, W. N., Robertson, J. T.

1975. Experiment I study on the genesis of cerebral vasospasm. *Stroke* 6:52-57
133. White, R., Terragno, D. A., Terragno, N. A., Hagen, A. A., Robertson, J. T. 1977. Prostaglandins in porcine cerebral blood vessels. *Stroke* 8:135
134. Wishaw, I. Q., Veale, W. L. 1974. Comparison of the effect of prostaglandin E₁ and norepinephrine injected into the brain on ingestive behaviour in the rat. *Pharmacol. Biochem. Behav.* 2:421-25
135. Wolfe, L. S. 1975. Possible roles of prostaglandins in the nervous system. In *Advances in Neurochemistry*, ed. M. Aprison, B. W. Agranoff, 1:1-49. New York: Plenum. 309 pp.
136. Wolfe, L. S. 1978. Some facts and thoughts on the biosynthesis of prostaglandins and thromboxanes in brain. See Ref. 27, pp. 215-20
137. Wolfe, L. S., Mamer, O. A. 1974. Measurement of prostaglandin F_{2a} levels in human cerebrospinal fluid in normal and pathological conditions. *Prostaglandins* 9:183-92
138. Woolf, C. J., Willies, G. H., Rosendorff, C. 1977. Pyrogen, prostaglandin and cyclic AMP fevers in the rabbit. In *Drugs, Biogenic Amines and Body Temperature*, ed. K. E. Cooper, P. Lom x, E. Schönbaum, pp. 136-39. Basel: Karger. 283 pp.
139. Yamamoto, Y. L., Feindel, W., Wolfe, L. S., Katoh, H., Hodge, C. P. 1972. Experimental vasoconstriction of cerebral arteries by prostaglandins. *J. Neurosurg.* 37:385-97
140. Ziel, R., Krupp, P. 1976. Influence of endogenous pyrogen on the cerebral prostaglandin-synthetase system. *Experientia* 32:1451-53